



β hydroxy butyrate Protocol

Version: 1

Replaced by version

Edited by: Peter Havel - UC Davis Metabolism and Endocrinology Core

[Summary](#)

[Reagents and Materials](#)

[Protocol](#)

[Reagent Preparation](#)

Summary:

When a sample is mixed with R1, AcAc in the sample is broken down to acetone by AADC. Upon addition of R2, 3-HB in the sample is oxidized in the presence of 3-HBDH and Thio-NAD. This oxidation triggers the cyclic reactions. Since the original AcAc in the sample has been removed, only 3-HB is assayed by measuring the rate of Thio-NADH production spectrophotometrically.

Reagents and Materials:

Reagent/Material	Vendor	Stock Number
Calibrator	Wako	412-73791
Reagents	Wako	417-73501 413-73601
Microplate		
Platereader		

Protocol:

1. Reconstitute R1 and R2 using the buffers provided.
2. Add 4 μ l of calibrator and sample to each well.
3. Add 270 μ l of R1 to each well. Incubate at 37°C for 5 minutes.

IMPORTANT: Make sure not to add any bubbles to the wells when dispensing reagents, this will interfere with reading in the platereader.

4. Add 90 μ l of R2 to each well. Incubate at 37°C for 2 minutes. Read at 405 nm. Then continue reading every 30 seconds for 2 minutes.

5. Calculate the slope of the reaction for each well. The assay will be linear so the unknown samples can be calculated as $(\text{Sample } \Delta\text{OD}/\text{min} \div \text{Calibrator } \Delta\text{OD}/\text{min}) \times \text{Calibrator Concentration}$.

Reagent Preparation:

R1 – reconstitute with buffer provided

R2 – reconstitute with buffer provided